Directory	Sites	Ligands	Method	Notes
Cyp_CsA+	1	1	fluorescence intensity	Basic protein-ligand Kd determination  Cyp = recombinant human cyclophilin  CsA+ = Cyclosporin A labeled with a fluorescent probe (+)  This is the control experiment, a prerequisite for analysis of competitive displacement assay
Cyp_CsA_CsA+	1	2	fluorescence intensity	data using the unlabeled CsA (molecule of interest)  Competitive ligand displacement assay  Cyp = recombinant human cyclophilin  CsA = Cyclosporin A  CsA+ = Cyclosporin A labeled with a fluorescent probe (+)  Kd for the spectroscopically "invisible" ligand is determined by a competing out the "visible"
RIZ1_H3pep	1	1	NMR	ligand from the protein-ligand complex. Compare two methods of analysis: (1) determine Kd for CsA+ separately. or (2) determine both Kd's simultaneously.  NMR chemical shift titration to determine weak protein-protein Kd  RIZ1 = tumor supporesor protein  H3pep = synthetic peptide representing histone H3
TDG_16U11+	2	1	fluorescence polarization	Combine chemical shifts for multiple nuclei ( <sup>1</sup> H and/or <sup>15</sup> H) to increase overall sensitivity of Kd measurement. <b>Two binding sites: one strong ("specific") and one weak ("nonspecific") TDG</b> = thymine DNA glycosylase <b>16U11+</b> = Texas Red (+) labeled 28 bp DNA fragment, U as target nucleotide
ThT_22AG	4	1	ESI-MS	Two-site vs. one-site model discrimination analysis.  Mass spectrometry titration: Four cooperative binding sites  ThT = Thioflavin T  22AG = human telomeric G-quadruplex sequence dAGGG(TTAGGG) <sub>3</sub>
WIN61651_p56lck	2	2	<sup>32</sup> P liquid scintillation counting	"Progressive cooperativity": Each added ligand is causing the subsequent ligand to bind ever more strongly (up to four binding sites).  Initial rate enzyme kinetics: determine mechanism / mode of inhibition  WIN61651 = small molecule kinase inhibitor (Sterling Winthrop)  p56lck = p56lck kinase
Lig_Prot_BDom	1	2	(unspecified)	Determining the mode of inhibition (competitive, non-competitive, mixed type) is strongly affected by substrate inhibition.  Full-length protein and just the binding domain compete for a ligand of interest  Lig = some type of ligand (undisclosed)  Prot = some type of protein (undisclosed)
				BDom = a binding domain taken out of this protein and expressed separately  The purpose of this experiement is to determine the affinity (Kd) between the binding domain and the ligand. One big problem is that only the binding to the full-length protein can be observed, and most importantly the binding to the full length protein is approximately 50 times weaker than binding to the binding domain. This very large difference in binding affinities presents a significant challenge for experiment design.